

Title:

Authors:

DOI:

Date submitted:

Date accepted:

Available online:

Citation:

Special issue:

IMPORTANT—This version of an accepted manuscript that will appear in *Bulletin of Marine Science* is a submission that has been peer reviewed and accepted for publication; however, it has not yet been edited and/or formatted, and items that will appear in the final publication may be missing. During the article production process, the text and other elements of this article are likely to change.

When the final article is available, this FastTrack article will be removed and the final version will replace it. The date the article was first made available online will be carried over, but the year of publication will correspond to the volume/issue of inclusion.

Accepted manuscripts can be cited in text as “In Press”.

Efficacy of probability surveys for ESA-listed coral species in southern Florida.

Dione W. Swanson^{1,2*}, Steven G. Smith^{1,2}, T. Shay Viehman³, Steven L. Miller⁴, Jennifer Stein⁵,
Rob Ruzicka⁵, Jennifer C. Samson⁶

¹ Cooperative Institute for Marine and Atmospheric Studies, University of Miami Rosenstiel
School of Marine, Atmospheric, and Earth Science, Miami, Florida, United States

² NOAA National Marine Fisheries Service, Southeast Fisheries Science Center, Miami, Florida,
United States

³ NOAA National Ocean Service, National Centers for Coastal Ocean Science, Beaufort, North
Carolina, United States

⁴ Nova Southeastern University, Fort Lauderdale, Florida, United States

⁵ Florida Fish and Wildlife Research Institute, St. Petersburg, Florida, United States

⁶ NOAA National Marine Fisheries Service, Pacific Islands Fisheries Science Center, Honolulu,
Hawaii, United States

*Corresponding Author Email: <d.swanson1@umiami.edu>

Running Title: Probability surveys for ESA-listed coral species

Keywords: Survey design, ESA-listed coral species

ABSTRACT

As coral populations have declined on coral reefs in the United States, one federal management tool to conserve species is the U.S. Endangered Species Act (ESA). Evaluation of the population status and trends for each species at the ecosystem scale requires periodic surveys, but surveys require a sampling effort sufficient to detect change at high levels of precision over time. Therefore, we assessed the feasibility of diver-based survey data to quantitatively evaluate the seven ESA-listed coral species on Florida's coral reefs. Data are from two complementary monitoring programs constructed from the same probability-based survey design that incorporates stratification created from density mean-variance spatial patterns of abundant target species. The survey design also included along-shore sub-regional gradients and marine spatial planning to tailor the sample frame specifically to the spatial distributions of each threatened species. We evaluated a multi-species design by comparing precision-sample size relationships from two stratification schemes. Results showed that constructing a composite scheme from the habitat use of 4 ESA-listed species resulted in a multi-species design with predicted precision estimates (CV) from 7.5% to 20% for six threatened species. These precision levels enable detection of change in mean density from 15% to 40% (approximately 2 SEs or twice the CV). This level of precision ranked as high as those for many abundant species for four threatened species and somewhat lower for the two rarest species. However, controlling survey precision at the regional scale would likely require sample sizes at least 3 times larger than a typical survey effort for most ESA-listed species.

INTRODUCTION

Coral declines over the last several decades are well-documented (Hughes and Tanner 2000, Gardner et al. 2003, Bruno and Selig 2007, Knowlton and Jackson 2008, Dietzel et al. 2020, Tebbett et al. 2023) and the causes are clear. Global warming causes coral bleaching (Glynn 1993, Hoegh-Guldberg 1999, Hughes et al. 2018), and coral diseases are widespread and increasing in severity (Bruno et al. 2007, Walton et al. 2018, Muller et al. 2020). Hurricanes cause damage at local and regional scales (Gardner et al. 2005, Madden et al. 2023). Geography is important in places like Florida where winter cold fronts have caused periodic and large-scale mortality (Lirman et al. 2011). Globally, coral cover has declined from $36\pm1.4\%$ to $19\pm0.4\%$ from 1997 to 2018 (47% decline; Tebbett et al. 2023). Depending on the region and habitat type, coral cover in Florida has declined approximately 12-76% from the 1970s to 2021 (Somerfield et al. 2008, Ruzicka et al. 2013, Viehman et al. 2023). In comparison, coral cover on high relief reefs in the US Virgin Islands has declined between 58-63% from 1999 to 2021 (Viehman et al. 2023). These changes in coral cover translate to decreases in colony abundance and changes to the demographic structure of individual coral species (Dietzel et al. 2020). For example, many of the abundant, reef building and ecologically important species in Florida are now considered occasional or rare (Porter and Meier 1992, Ruzicka et al. 2013) and the iconic pillar coral, *Dendrogyra cylindrus*, is regionally extinct (Jones et al. 2021, Neely et al. 2021).

In response to the decline in population abundance of corals, assessments of species' extinction risk have been conducted globally for the International Union for Conservation of Nature (IUCN) Red List of Threatened Species and regionally, in the United States, through the U.S. Endangered Species Act listing process with the intention to manage and conserve declining species (Mace et al. 2008, Federal Register 2006, 2014, Gutierrez et al. 2024). The criteria of

listing threatened species by both processes contain similar evaluation elements such as the magnitude of population abundance decline and changes in productivity and spatial structure (Brainard et al. 2011, IUCN 2012). However, a majority of the data used in these risk assessments are acquired from surveys designed to evaluate coral community and ecosystem function indicators not coral populations (Mace et al. 2008, Brainard et al. 2011, Gutierrez et al. 2024). For the survey data that are designed to evaluate coral populations, the selected target species are usually not the threatened IUCN Red List and ESA-listed species (NMFS 2022).

Seven reef-building coral species are listed as threatened under the ESA in Florida and the Caribbean – *Acropora cervicornis*, *A. palmata*, *Orbicella annularis*, *O. faveolata*, *O. franksi*, *Dendrogyra cylindrus*, and *Mycetophyllia ferox* (Federal Register 2006, 2014). The recovery documents developed for these listed species identify the biological, conservation, and threat assessments as well as the actions needed to recover the species (NMFS 2015a, b; NMFS 2022). Biological assessment actions include improving the understanding of population demographic structure, genetic structure, and population dynamics and include implementation of monitoring species abundance and distribution over time to evaluate population status (NMFS 2015a, b). For each species, effective monitoring requires periodic surveys to estimate population abundance at the ecosystem scale with a sampling effort sufficient to detect change at high levels of precision over time. While U.S. coral reef jurisdictions conduct repeated, regular ecosystem-scale coral surveys (NOAA CRCP 2021), and Florida has additional monitoring specific to coral stressors (i.e., bleaching; Waddell 2017), neither of these surveys have been specifically designed to provide population estimates for ESA-listed species with low abundance.

A probabilistic sampling approach can provide the underlying framework for a survey design that is specifically tailored to these threatened species. For Florida's coral reefs, two

complementary programs (NOAA's National Coral Reef Monitoring Program (NCRMP) and Florida Reef Resiliency Program's Disturbance Response Monitoring (DRM)) use a single survey design that optimizes sampling efficiency to provide high-precision estimates of population metrics at relatively low sample sizes (Miller et al. 2002, Smith et al. 2011a). The stratified random survey design reduces population variance based on two factors. First, coral density varies based on different reef habitat features such as depth, rugosity, and reef type (e.g. hardbottom, spur-and-groove, patch reef). Second, habitats with high density typically also have high variance, while habitats with low density have low variance. Benthic maps of reef habitat features are incorporated into the stratification of the survey area (i.e. domain or sample frame). Stratification partitions the survey domain into sectors of low, moderate, and high variance of coral density. The optimization process allocates sampling (the number of sites to be sampled) by stratum area and the variance of target species density. The survey design of the two Florida programs is optimized for a select number of coral species that are abundant and ecologically important. Both programs, however, include all observed coral species in field surveys. The long-term monitoring data provided by these complementary programs allows for the statistical evaluation of survey design parameters needed to optimize surveys specific to rare ESA-listed species. Even though tailoring a survey design through stratification and optimal allocation is standard practice for rare species (Lohr 2022), there remains the potential of not observing these species when they are actually present in the sample frame (Chao and Lee 1992). Estimation of sample coverage provides a metric to evaluate if rare species are adequately sampled in a survey (Chiu 2023).

Given the need to monitor change in ESA-listed species and the challenge of low density, the goal of this study was to determine which survey design strategy is best suited to obtain precise

abundance estimates in a cost-effective manner. The objectives of our study were to: 1) evaluate sampling coverage, 2) evaluate the most effective survey design and obtainable levels of precision of future surveys for selected ESA-listed coral species using density as the design metric, 3) determine the most effective way to tailor the stratification for ESA species, and 4) evaluate survey effort requirements in relation to precision and spatial scale (e.g. domain and region estimates).

METHODS

Study Area

The study area encompassed reef habitats of Florida's coral reef ecosystem extending for 476 km from Broward County to the Dry Tortugas (Fig. 1), including three main geographic regions – Southeast Florida, Florida Keys, and Dry Tortugas. In the Southeast Florida region, reef habitat types are well-developed linear reef complexes, a series of ridges, patch reefs, and colonized pavement. The Florida Keys and Dry Tortugas regions consist of a semi-continuous series of bank-barrier reefs (i.e. fore reef) and the Florida Keys also includes patch reefs and back reef areas between the fore reef and either land or barrier islands extending from Biscayne National Park through the Florida Keys.

Diver Probability Surveys

This study utilized data from diver-based surveys of stony corals conducted by two ongoing long-term monitoring programs: 1) NOAA's National Coral Reef Monitoring Program (NCRMP), which surveys biennially to evaluate population status and change, and 2) Florida Reef Resiliency Program's Disturbance Response Monitoring (DRM), which surveys corals annually for bleaching and disease. The data in this study included the surveys from 2014 to

2016 for NCRMP and from 2013 to 2016 for DRM. Both programs use methods and stratified random design detailed in Smith et al. (2011a). The survey domain encompassed all mapped live coral habitat area from nearshore to offshore (0-33m depth for NCRMP; 0-20m depth for DRM) between Broward County and the Dry Tortugas. For both programs, stratified random site selection was based on a gridded sample frame consisting of 100 x 100 m grid cells (primary sampling units, PSU) and stratified by cross-shelf zonation (e.g., back reef, fore reef), depth, reef morphology (e.g., patch reefs, spur-groove reefs), and reef complexity. The number, size and relative condition of adult scleractinian coral colonies (maximum diameter ≥ 4 cm) for all coral species were measured in situ along one (NCRMP) or two (DRM) belt transect(s) (10m² each transect) per PSU (NCCOS 2017, DRM 2017). An individual colony was considered to be a continuous skeletal unit; therefore, a colony that was part of the same skeleton but divided into two or more separate pieces of live tissue was considered to be one colony (Meesters et al. 1996). For each PSU, density was computed as the number of colonies per m² and occurrence was scored as the presence (1) or absence (0) of a given species within the 10m² belt transect.

Sample coverage was estimated using the abundance-based sample coverage equation in Chiu (2023). Population metrics estimated from diver surveys included mean density ($\bar{D}$, number of colonies per unit area) and proportion occurrence ($\bar{P}$, proportion of PSUs with one or more colony). Estimation of strata- and domain-level population metrics followed standard procedures for stratified random sampling (Cochran 1977, Lohr 2022). Domain-level estimates were computed as the weighted average of stratum-level estimates, illustrated for mean density as

$$\bar{D}_{st} = \sum_h w_h \bar{D}_h \quad , \quad (1)$$

where $\bar{D}_{st}$ is mean colony density for the stratified domain (i.e., spatial sample frame), $\bar{D}_h$ is mean colony density in stratum h , and w_h is the stratum weighting factor, the stratum proportion of sample units in the domain. The weighting factor was computed as

$$w_h = \frac{N_h}{\sum_h N_h} \quad , \quad (2)$$

where N_h is the total possible stratum PSUs. In this formulation, the stratum weighting factor equals the stratum proportion of area within the sampling domain. Computation of the variance of domain-wide mean density ($var[\bar{D}_{st}]$) was computed in a similar fashion as the weighted average of strata variances,

$$var[\bar{D}_{st}] = \sum_h w_h^2 var[\bar{D}_h] \quad , \quad (3)$$

where the variance of mean density in stratum h ($var[\bar{D}_h]$) was,

$$var[\bar{D}_h] = \left(1 - \frac{n_h}{N_h}\right) \frac{s_h^2}{n_h} \quad , \quad (4)$$

and the sample variance (s_h^2) among PSUs i in stratum h was computed as,

$$s_h^2 = \frac{\sum_i (D_{hi} - \bar{D}_h)^2}{n_h - 1} \quad . \quad (5)$$

The quantity $\left(1 - \frac{n_h}{N_h}\right)$ in equation 4 is the finite population correction factor. Survey error, as a measure of precision, was estimated as the coefficient of variation of mean density,

$$CV(\bar{D}_{st}) = \frac{SE(\bar{D}_{st})}{\bar{D}_{st}} \quad , \quad (6)$$

the standard error expressed as a proportion of the mean, where $SE(\bar{D}_{st}) = \sqrt{var(\bar{D}_{st})}$. For the NCRMP and DRM surveys, sampling effort was allocated among strata according to a Neyman strategy (Cochran 1977), taking into account both stratum area (i.e., number of PSUs) and stratum variance of density for a suite of abundant coral species.

Design Performance Analysis for ESA Species

The NCRMP/DRM stratification-allocation scheme enabled estimation of domain-wide population metrics for target abundant coral species at substantially higher precision and lower cost (i.e., sample sizes) compared to a simple random design (Smith et al. 2011a). The feasibility of diver surveys to monitor less abundant populations of ESA corals was analyzed with respect to detecting changes over time (i.e., from survey to survey). Precision-sample size relationships were developed as in Smith et al. (2011a) using

$$n^* = \frac{(\sum_h w_h s_h)^2}{V(\bar{D}_{st}) + \frac{1}{N} \sum_h w_h s_h^2}, \quad (7)$$

where n^* is the number of sampled PSUs required to achieve a specified variance for a future survey, s_h is the standard deviation of density in stratum h , s_h^2 is the sample variance of density in stratum h , $V(\bar{D}_{st})$ is the desired variance for mean density, and N is the total number of possible PSUs in the domain. Estimation of n^* presumes that PSUs will be distributed among strata according to a Neyman allocation scheme,

$$n_h = n \left[\frac{w_h s_h}{\sum_h w_h s_h} \right], \quad (8)$$

which accounts for both stratum area (w_h) and variance (s_h). Desired variance of domain-wide mean density was estimated using a target CV,

$$V(\bar{D}_{st}) = (CV(\bar{D}_{st}) \cdot \bar{D}_{st})^2. \quad (9)$$

The 95% confidence interval for $\bar{D}_{st}$ is approximately two standard errors; thus, the minimum detectable change in mean density from one survey to the next is approximately twice the CV.

The NCRMP/DRM cross-shelf stratification scheme (Table 1) was based on density mean-variance spatial patterns for abundant coral species. These analysis strata (AS) were the basis for Neyman allocation (eq. 8), and estimation of population metrics (e.g., eq. 1) and associated standard errors. Investigations of stratification schemes more tailored to spatial distributions of ESA species took advantage of an inherent feature of the NCRMP/DRM surveys that incorporated additional factors (along-shore gradients, marine spatial planning) that may impact spatial distributions of corals. For random selection, the allocated PSUs to sample in each AS were first sub-allocated by geographical subregions (Fig. 1) and spatial management zones (marine reserves, fishing zones) proportional to their respective areas within a stratum. This proportional sub-allocation strategy to smaller ‘field strata’ (FS) was the practical equivalent of simple random sampling within an analysis stratum, and thus did not alter the original AS selection probability of PSUs. This provided flexibility for further dividing AS by subregions and/or management zones to construct more effective stratification schemes for ESA corals without introducing bias into variance estimates (Cochran 1977). Considering colony density as a measure of per unit habitat use (Manly et al. 1993), the building blocks of FS were used to construct habitat use strata (HUS) for ESA species that spatially partitioned mean and variance of density into low, moderate, and high use categories. This procedure entailed comparing individual FS means (e.g., subregions) with the corresponding AS mean (e.g., a habitat-depth

category), combining spatially adjacent FS with similar means into larger HUS, and then confirming the new groupings with statistical tests for differences in means and variances.

Design performance was analyzed for ESA coral species as well as two abundant species, *Montastraea cavernosa* and *Siderastrea siderea*, as references. Three stratification schemes were compared: (1) habitat use (HUS), (2) the NCRMP/DRM stratification (AS), and (3) simple random sampling (SRS). Analysis of SRS required a re-weighting of the NCRMP/DRM stratified random observations to meet the assumptions for simple random sampling, namely that sampling was conducted proportional to strata area. In contrast, the AS design employed by NCRMP/DRM used a Neyman scheme that allocated sampling effort according to both strata area and variance of density (eq. 8). The stratum sample size (n_h) condition for SRS is

$$\frac{n_{h,SRS}}{n} = \frac{N_h}{N} = w_h, \quad (10)$$

in which $n_{h,SRS}$ is proportional to N_h , the total PSUs in a stratum. Computation of $n_{h,SRS}$ was carried out for the survey AS design by

$$n_{h,SRS} = w_h \times n, \quad (11)$$

which was then used to compute the observation weight t in stratum h for a SRS design,

$$t_{h,SRS} = \frac{n_{h,SRS}}{n_{h,AS}}. \quad (12)$$

Evaluation of precision-sample size relationships for SRS entailed multiplying stratum weights $t_{h,SRS}$ by each observation in the respective analysis stratum h , and then computing n^* for a simple random design,

$$n^*(SRS) = \frac{s^2}{V(\bar{D}) + \frac{1}{N}s^2} \quad (13)$$

All computations were carried out using SAS (SAS Institute, v 9.4) and R (R Core Team, v3.1.3) software packages.

RESULTS

The 2013-2016 NCRMP/DRM surveys sampled 758 PSUs (Fig. 1, Table 1) and six ESA-listed coral species were observed: *A. cervicornis*, *A. palmata*, *M. ferox*, *O. annularis*, *O. faveolata*, and *O. franksi*. A seventh ESA coral, *Dendrogyra cylindrus*, is known to inhabit Florida's coral reefs, but is present at such low numbers that it was not observed in these surveys. The estimate of sample coverage for the survey was 0.999943. To aid interpretation of design performance analyses, a $CV(\bar{D}_{st})$ of 15% was considered as a general benchmark for survey precision, and the sample size for the 2016 NCRMP/DRM survey of $n=451$ PSUs served as a reference point for a feasible level of sampling effort in a single survey year in Florida. The AS scheme employed by the NCRMP/DRM surveys was projected to require sample sizes less than 451 to achieve a 15% CV for three ESA species, and greater than 451 for three species (Fig. 2). For all six ESA species, the projected $n^*(15\%)$ for simple random sampling ranged from 3 to 10 times higher compared to the respective $n^*(15\%)$ for the AS design.

Construction of species-specific habitat use strata (HUS) that incorporated components of reef habitats such as cross-shelf zonation (e.g., back reef, fore reef), depth, reef morphology (e.g., patch reefs, spur-groove reefs), substrate complexity-rugosity, along shelf gradients, and marine spatial planning is illustrated for *O. faveolata* for the Florida Keys region using patch reef

habitats as the example (Fig. 3). Mean density in patch reef habitats differed by cross-shelf analysis strata (inshore, mid-channel, offshore), and thus the original AS scheme was left unchanged (Fig. 3A). Within the mid-channel patch reef AS, mean density in the lower Keys subregion was similar to the Keys-wide average, mean densities in the adjacent middle and upper Keys subregions were above average, and mean density in the Biscayne subregion was below average (Fig. 3B). The mid-channel patch reef AS was subsequently divided into habitat use strata corresponding to three subregion groupings: lower Keys, middle-upper Keys, and Biscayne. The lower Keys mid-channel patch reefs were further subdivided into smaller HUS by management zone (open to fishing, marine protected areas), based on differences in mean density (Fig. 3C). This procedure resulted in 28 HUS for *O. faveolata* for the southern Florida survey domain (Table 2), compared to the original 30 analysis strata (Table 1). The number of HUS for three additional ESA species (*A. cervicornis*, *O. annularis*, *O. franksi*) ranged from 12 to 26 (Table 2). Comparing precision-cost relationships between the AS and species-specific HUS schemes (Fig. 2), $n^*(15\%)$ was lower for HUS for *A. cervicornis*, slightly higher for *O. franksi*, and similar to AS for *O. annularis* and *O. faveolata*. Two species, *A. palmata* and *M. ferox*, were too rare for habitat use analysis.

To evaluate a multispecies design, combining the HUS schemes for the four ESA species analyzed resulted in 55 unique strata (Table 2). Precision-sample size relationships were compared for this composite species habitat use stratification with the AS scheme for the six ESA species and two abundant coral species (Table 3). For the AS stratification (Table 3A), the expected precision for the benchmark annual survey effort ($n=451$) ranged from about 4% CV for the most abundant species (*S. siderea*: occupancy rate $\bar{P} \approx 0.85$, $\bar{D} \approx 1.3$ colonies per m^2) to about 30% CV for the two rare species (*A. palmata* and *M. ferox*: $\bar{P} \leq 0.01$, $\bar{D} \approx 0.001$).

However, the composite species HUS scheme improved the expected precision for all species (Table 3B): the projected CVs were 5% or less for *M. cavernosa* and *S. siderea* ($\bar{P} > 0.60$), about 7.5% for *O. faveolata* and *O. franksi* ($\bar{P} \approx 0.10$ to 0.30), about 15% for *A. cervicornis* and *O. annularis* ($\bar{P} = 0.04$), and about 20% for the two rare species. Evaluating the composite HUS scheme at the region scale (Dry Tortugas, Florida Keys, Broward-Miami) indicated that projected overall sample sizes would be considerably higher to achieve a precision of 15% for separate surveys in each region compared to a southern Florida survey domain encompassing all three regions (Table 4).

DISCUSSION

Combined diver probability surveys from NCRMP and DRM in southern Florida had a high sample coverage estimate and therefore provided the foundation to quantitatively evaluate the feasibility of surveys to monitor ESA-listed coral species. These surveys incorporate stratification based on density mean-variance spatial patterns of abundant target species but are structured to include additional factors (along-shore gradients, marine spatial planning) that may impact spatial distributions of corals. These fine-scale elements were the building blocks to tailor the sample frame to spatial distributions of each threatened species. The NCRMP/DRM 2016 survey effort was used as a benchmark of a reasonable sample size for a future ESA-listed species survey. Comparison of precision-sample size relationships from two stratification schemes were conducted for a multi-species design. The results showed that constructing a composite scheme from the habitat use of 4 ESA species resulted in a multi-species design with predicted precision estimates (CV) ranging from 7.5% to 20% for all six species. These precision levels enable detection of change in mean density ranging from 15% to 40% (approximately 2

SEs or twice the CV). This level of precision for four threatened species ranked as high as those for many abundant species and somewhat lower for the two rarest species (Smith et al. 2011a). However, controlling survey precision at the regional scale (Broward-Miami, Florida Keys, and Dry Tortugas separately) would require sample sizes at least 3 times higher than the 2016 NCRMP/DRM survey effort for most ESA-listed species.

A key challenge in designing a survey for rare species is adequately detecting these species when they are actually present. The pooled survey data in this study was optimally allocated for target species that did not include the ESA-listed species which necessitated an estimate of sample coverage. Sample coverage is a metric of sample completeness and is defined as the proportion of individuals of each species in a given sample where values range from 0 to 1 (Chao and Lee 1992, Chiu 2023). Our estimate of sample coverage was close to 1 (0.999943) which signified that we were able to adequately observe the six ESA-listed coral species included in this study. However, the survey did not detect *Dendrogyra cylindrus*. This species has been considered naturally rare on the Florida reef tract (Chan et al. 2019). The number of known *Dendrogyra* colonies in the Florida population (SE Florida, Florida Keys, and Dry Tortugas) was 819 individuals from 2014 to 2020 (Neely et al. 2021). However, after fate tracking all colonies during this time period, the population experienced a 93% loss of colonies and the species was determined to be functionally extinct (Neely et al. 2021).

This study examined both the stratification and allocation components of survey design for ESA-listed coral species by comparing the precision-sample size relationships among stratification schemes. As expected, the two stratified designs (AS and HUS) provided a higher level of precision at much less effort than simple random design for individual species. Given the heterogeneous nature of both habitat and the distribution of abundance, stratification effectively

created different subareas of homogeneous abundance and variance. Using a single-species design with the existing NCRMP/DRM stratification scheme (AS) or the species-specific habitat use scheme (HUS), the predicted sampling effort required to achieve a 15% CV was less than the benchmark effort (451 PSUs) for the three *Orbicella* species. A predicted CV of less than 15% at a typical level of sampling effort would detect a minimum 30% change in density. Although the predicted sampling effort required to achieve a 15% CV was higher than 451 PSUs for *A. cervicornis* for both the AS and HUS scheme, the precision-sample size relationship (Figure 2C) indicated a CV of less than 20% at a sample size of 451 which would detect an approximate 40% change in density or 0.0058 colonies/m². For *A. palmata* and *M. ferox*, the sampling effort needed to achieve a 15% CV was more than 3 times higher than the benchmark effort. Overall, single-species stratified designs using the NCRMP/DRM stratification-allocation scheme would provide reasonable change detection in density for at least four species.

Surveys are frequently designed using multiple target species to address programmatic priorities and maximize resources (e.g. Smith et al. 2011a, b). Ideally, a survey design targeting the six species in this study would be the most cost effective compared to individual surveys for each species. We used precision-sample size relationships to compare a composite HUS from four species to the AS to determine if it would improve performance of a future multi-species survey for the six ESA-listed species. The composite HUS improved expected precision for all six species. A predicted CV of less than 7.5% at a typical level of sampling effort would detect a minimum 15% change in density for *O. faveolata* and *O. franksi*, or a density change of ± 0.0111 colonies/m² and ± 0.0056 colonies/m² respectively. For *A. cervicornis* and *O. annularis*, the predicted CV of less than 15% would detect a minimum of 30% change in density or ± 0.0062 colonies/m² for *A. cervicornis* and ± 0.0041 colonies/m² for *O. annularis*. The predicted precision

for the two rarest species, *M. ferox* and *A. palmata*, was 20% resulting in a 40% change in density or ± 0.005 colonies/m² and ± 0.0002 colonies/m² respectively. The improvement in expected precision for all species including the two rarest species (*A. palmata* and *M. ferox*) using the composite HUS compared to AS showed that a stratification scheme based on spatial distribution of abundance and variance of these threatened species is effective for detecting change in population density at a feasible level of sampling effort in a single survey. The composite HUS also performed better than the species-specific HUS schemes for all four species.

Two abundant species were included in the comparison of the composite HUS and AS schemes to evaluate how a survey designed for ESA-listed species would affect their performance in terms of predicted CV. The predicted precision for these species also improved with the composite HUS scheme. There was a reduction in predicted sample size for *S. siderea* and the CV decreased from approximately 7.5% to less than 5% for *M. cavernosa*. Given the benchmark sampling effort, the composite HUS designed for ESA-listed species could be implemented in the current NCRMP/DRM surveys and maintain high levels of precision for abundant species. The NCRMP reef fish surveys use an analogous strategy and target exploited, less abundant fish species while achieving high levels of precision for abundant species (Smith et al 2011b). This strategy would benefit both the primary goals of the NCRMP and DRM surveys and the ESA recovery evaluation needs.

The composite HUS precision-sample size relationship demonstrates two important aspects of survey design – sample variance with respect to density and practical minimum sample size. Projection of sampling effort at different levels of precision (Table 3B) showed a distinct pattern with the abundance rank of each species. At low levels of population density, the precision was

low and thus more samples are required. As the density increases, precision stabilizes, and fewer samples are needed to obtain a precise estimate. Also, the projected sample size (n^*) values for *S. siderea* and *M. cavernosa* become exceedingly small as % CV increases (Table 3B). The projected sample size is a theoretical projection and can produce low sample sizes for the most abundant species. The practical minimum sample size for a survey is twice the number of strata to ensure that each stratum has a minimum number of 2 samples. The practical minimum sample size was 60 and 110 for AS and HUS respectively.

The survey data in this study combined three regions in southern Florida. If a region-specific estimate of ESA-listed species is desired, our results showed that sampling to control for precision of density estimates at the regional scale using the composite HUS would require considerably more sampling effort than the sample size needed to control the same level of precision of the entire domain encompassing all three regions (Table 4). For the least abundant species, the sampling effort needed at the regional scale was much higher. Similar results have been found for reef fish species, highlighting the importance of clearly defined objectives for probability-based surveys with respect to target species and management priorities (Smith et al. 2011b, Bryan et al 2016).

Following the listing of all seven ESA-listed species, population status reviews are conducted every five years (NMFS 2020). In Florida, the NCRMP and DRM survey estimates have been incorporated into the most recent population status evaluation of these species along with data from permanent monitoring sites (e.g. Ruzicka et al. 2013) and studies that have investigated genetic structure, reproductive/fertilization success and population dynamic rates of some ESA-listed species (e.g. Kabay 2016, Miller et al. 2016a,b; Williams et al. 2017; Neely et al. 2018; Neely et al. 2020; Williams et al. 2020, Neely et al. 2021). Population and stratum-specific

abundance estimates from the NCRMP and DRM surveys serve as a complement to smaller scale studies where localized changes can be placed in context of the larger scale surveys and the population demographic structure estimates could improve predictive modeling efforts.

Collectively, these surveys and specific studies address the biological actions needed to assess recovery and improve the evaluation of ESA-listed species. Here we show that moderate adjustments to the current survey design in Florida can increase the precision of change detection in populations of six ESA-listed coral species while retaining the detection power in more abundant species. This further optimization of sampling design would enhance understanding of the ecosystem, better inform management decisions, and decrease the cost of evaluating species of interest.

Probability surveys incorporate a key iterative learning component as standard practice that keeps the design tailored to the changing dynamics of a species. Practically, this means that when planning each survey using the stratification scheme for these rare species, survey performance is evaluated and an optimal (Neyman) allocation of sampling effort is conducted using the three most recent years of survey data. This provides a mechanism for iterative optimization of survey efficiency under changing environmental conditions that potentially impact these species.

ACKNOWLEDGEMENTS

The authors greatly acknowledge the efforts of the contributors who designed surveys, collected data and provided logistical support on the Florida Reef tract as part of NOAA's National Coral Reef Monitoring Program (NCRMP) and Florida Reef Resiliency Program's Disturbance Response Monitoring (DRM). NCRMP funding is provided by NOAA's Coral Reef Conservation Program project 743.

This study was part of a larger project entitled 'A tool to assess the population status of [threatened] coral species in U.S. waters' funded by NOAA Office of Science and Technology's National Protected Species Toolbox Initiative:

<https://www.st.nmfs.noaa.gov/protected-species-science/Science-Communication-and-Coordination/toolbox>.

This research was carried out [in part] under the auspices of the Cooperative Institute for Marine and Atmospheric Studies (CIMAS), a Cooperative Institute of the University of Miami and the National Oceanic and Atmospheric Administration, cooperative agreement #NA20OAR4320472.

LITERATURE CITED

- Brainard RE, Birkeland C, Eakin CM, McElhany P, Miller MW, Patterson M, Piniak GA. 2011. Status review report of 82 candidate coral species petitioned under the U.S. Endangered Species Act. U.S. Dep. Commer., NOAA Tech. Memo., NOAA-TM-NMFS-PIFSC-27, 530 p. +1 Appendix.
- Bruno JF, Selig ER. 2007. Regional decline of coral cover in the Indo-Pacific: Timing, Extent, and Subregional Comparisons. *PLOS ONE* 8:e711.
- Bryan DR, Smith SG, Ault JS, Feeley MW, Menza CW. 2016. Feasibility of a region-wide probability survey for coral reef fish in Puerto Rico and the U.S. Virgin Islands. *Mar Coast Fish* 8(1):135–146. <https://doi.org/10.1080/19425120.2015.1082520>
- Chao, A., & Lee, S. M. (1992). Estimating the number of classes via sample coverage. *J Am Stat Assoc* 87(417), 210-217.
- Chiu, C. H. (2023). Sample coverage estimation, rarefaction, and extrapolation based on sample-based abundance data. *Ecology*, 104(8), e4099.
- Cochran WG. 1977. Sampling techniques. New York, John Wiley and Sons.
- Dietzel A, Bode M, Connolly SR, Hughes TP. 2020. Long-term shifts in the colony size structure of coral populations along the Great Barrier Reef. *Proc. R. Soc. B* 287: 20201432. <http://dx.doi.org/10.1098/rspb.2020.1432>
- DRM. (2017). Florida Reef Resilience Program Disturbance Response Monitoring “DRM”. <https://www.ocean.floridamarine.org/FRRP>.
- Federal Register. 2006. Endangered and Threatened Species: Final Listing Determinations for Elkhorn and Staghorn Coral. *Federal Register* 71: 26852-27872.
- Federal Register. 2014. Endangered and Threatened Wildlife and Plants: Final Listing Determinations on Proposal To List 66 Reef-Building Coral Species and To Reclassify Elkhorn and Staghorn Corals. *Federal Register* 79:53852-54123.
- Florida Reef Resiliency Program – Disturbance Response Monitoring. 2017. FRRP-DRM In Water Protocol. <http://frfp.org/wp-content/uploads/2017/06/FRRP-DRM-In-Water-Protocol.pdf>.
- Gardner TA, Cote IM, Gill JA, Grant A, Watkinson AR. 2003. Long-term region-wide decline in Caribbean corals. *Science* 301: 958-960.
- Gardner TA, Cote IM, Gill JA, Grant A, Watkinson AR. 2005. Hurricanes and Caribbean coral reefs: impacts, recovery, patterns, and role in long-term decline. *Ecology* 86(1): 74-184.
- Glynn PW. 1993. Coral reef bleaching: ecological perspectives. *Coral Reefs* 12: 1-17.
- Gutierrez L, Polidoro B, Obura D, Cabada-Blanco F, Linardich C, Pettersson E, Pearce-Kelly P, Kemppinen K, Alvarado JJ, Alvarez-Filip L, et al. 2024. Half of Atlantic reef-building corals at elevated risk of extinction due to climate change and other threats. *PLoS ONE* 19(11): e0309354. <https://doi.org/10.1371/journal.pone.0309354>
- Hoegh-Guldberg O. 1999. Climate change, coral bleaching, and the future of the world’s coral reefs. *Mar Freshwater Res* 50: 839-866.
- Hughes TP, Kerry JT, Baird AH, Connolly SR, Dietzel A, Eakin CM, Heron SF, Hoey AS, Hoogenboom MO, Liu G, McWilliam MJ, Pears RJ, Pratchett MS, Skirving WJ, Stella JS, Torda G. 2018. Global warming transforms coral reef assemblages. *Nature* 556(7702): 492-496.
- Kabay L. 2016. Population demographics and sexual reproduction potential of the pillar coral, *Dendrogyra cylindrus*, on the Florida Reef Tract. Masters. Nova Southeastern University, Ft.

- Lauderdale, FL.
- Knowlton N, Jackson JBC. 2008. Shifting baselines, local impacts, and global change on coral reefs. *PLOS Biology*. 6(2):e54. <https://doi.org/10.1371/journal.pbio.0060054>
- Lirman D, Schopmeyer S, Manzello D, Gramer LJ, Precht WF, Muller-Karger F, Banks K, Barnes B, Bartels E, Bourque A et al. 2011. Severe 2010 cold-water event caused unprecedented mortality to corals of the Florida Reef Tract and reversed previous survivorship patterns. *PLoS ONE*. 6(8):e23047. <https://doi.org/10.1371/journal.pone.0023047>
- Lohr SL. 2022. Sampling Design and Analysis. 3rd ed. CRC Press, Boca Raton.
- Mace GM, Collar NJ, Gaston KJ, Hilton-Taylor C, Akçakaya HR, Leader-Williams N, Milner-Gulland EJ, Stuart SN. 2008. Quantification of Extinction Risk: ICUN's System for Classifying Threatened Species. *Conserv Biol*, 22: 1424-1442. <https://doi.org/10.1111/j.1523-1739.2008.01044.x>
- Madden IA, Mariwala A, Lindhart M, Narayan S, Arkema KK, Beck MW, Baker JW, Suckale J. 2023. Quantifying the fragility of coral reefs to hurricane impacts: a case study of the Florida Keys and Puerto Rico. *Environm Res Lett*. 18(2):024034. <https://doi.org/10.1088/1748-9326/acb451>
- Manly BFJ, McDonald LL, Thomas LL. 1993. Resource selection by animals. Chapman and Hall, London.
- Meesters EH, Wesseling I, Bak RPM. 1996. Partial mortality in three species of reef-building corals and the relation with colony morphology. *Bull Mar Sci* 58:838-852
- Miller MW, Kerr K, Williams DE. 2016a. Reef-scale trends in Florida *Acropora* spp. abundance and the effects of population enhancement. *PeerJ* 4:e2523.
- Miller MW, Williams DE, Fisch J. 2016b. Genet-specific spawning patterns in *Acropora palmata*. *Coral Reefs* 35:4. <https://doi.org/10.1007/s00338-016-1472-6>
- Miller SL, Swanson DW, Chiappone M. 2002. Multiple spatial scale assessment of coral reef and hardbottom community structure in the Florida Keys National Marine Sanctuary. *Proc 9th Intl Coral Reef Symp, Bali*, 1: 69-77.
- Muller EM, Sartor C, Alcaraz NI, van Woesik R. 2020. Spatial epidemiology of the Stony Coral Tissue Loss Disease in Florida. *Front Mar Sci*. 7. <https://doi.org/10.3389/fmars.2020.00163>
- National Centers for Coastal Ocean Science (NCCOS). 2017. NCRMP Caribbean-Gulf of Mexico Field Protocols. <https://coastalscience.noaa.gov/project/national-coral-reef-monitoring-program-biological-socioeconomic/>
- National Marine Fisheries Service. 2015a. Recovery Plan for Elkhorn (*Acropora palmata*) and Staghorn (*A. cervicornis*) Corals. Prepared by the *Acropora* Recovery Team for the National Marine Fisheries Service, Silver Spring, Maryland.
- National Marine Fisheries Service. 2015b. Recovery Outline: Pillar Coral, Rough Cactus Coral, Lobed Star Coral, Mountainous Star Coral, Boulder Star Coral. NMFS Southeast Regional Office. St. Petersburg, FL. <https://www.fisheries.noaa.gov/resource/document/5-caribbean-coral-species-recovery-outline>
- National Marine Fisheries Service. 2022. Staghorn coral (*Acropora cervicornis*), Elkhorn coral (*Acropora palmata*), Lobed star coral (*Orbicella annularis*), Mountainous star coral (*Orbicella faveolata*), Boulder star coral (*Orbicella franksi*), Rough cactus coral (*Mycetophyllia ferox*), Pillar coral (*Dendrogyra cylindrus*) 5-Year Review: Summary and Evaluation. NMFS Southeast Regional Office, St. Petersburg, FL. <https://doi.org/10.25923/jyer-dv55>

- Neely KL, Lewis CL, Lunz KS, Kabay L. 2018. Hermaphroditic spawning by the gonochoric pillar coral *Dendrogyra cylindrus* along the Florida reef tract. *Front Mar Sci* 8:434.
- Neely KL, Lewis CL, Macaulay KA. 2020. Disparities in spawning times between in situ and ex situ pillar corals. *Front Mar Sci* 7:643 <https://doi.org/10.3389/fmars.2020.00643>
- Neely KL, Lewis CL, Lunz KS, Kabay L. 2021. Rapid population decline of the pillar coral *Dendrogyra cylindrus* along the Florida reef tract. *Front Mar Sci* 8:656515 <https://doi.org/10.3389/fmars.2021.656515>
- NOAA Coral Reef Conservation Program. 2021. National Coral Reef Monitoring Plan. Silver Spring, MD: NOAA. <https://doi.org/10.25923/fqkq-w497>
- Porter JW, Meier OW. 1992. Quantification of loss and change in Floridian reef coral populations. *Am Zool.* 32(625-640).
- Ruzicka RR, Colella MA, Porter JW, Morrison JM, Kidney JA, Brinkhuis V, Lunz KS, Macaulay KA, Bartlett LA, Meyers MK, Colee J. 2013. Temporal changes in benthic assemblages on Florida Keys reefs 11 years after the 1997/1998 El Nino. *Mar Ecol Prog Ser* 489: 125-141.
- Smith SG, Ault JS, Bohnsack JA, Harper DE, Luo J, McClellan DB. 2011b. Multispecies survey design for assessing reef-fish stocks, spatially explicit management performance, and ecosystem condition. *Fish Res* 109(1):25-41.
- Smith SG, Swanson DW, Chiappone M, Miller SL, Ault JS. 2011a. Probability sampling of stony coral populations in the Florida Keys. *Environ Monit Assess.* 183(1-4): 121-138.
- Somerfield PJ, Jaap WC, Clarke KR, Callahan M, Hackett K, Porter J, Lybolt M, Tsokos C, Yanev G. 2008. Changes in coral reef communities among the Florida Keys, 1996-2003. *Coral Reefs* 27: 951-965.
- Tebbett SB, Connolly SR, Bellwood DR. 2023. Benthic composition changes on coral reefs at global scales. *Nature Ecology & Evolution* 7:71-81. <https://doi.org/10.1038/s41559-022-01937-2>
- Viehman TS, Groves SH, Grove LJW, Smith SG, Mudge L, Donovan C, Edwards K, Towle EK. 2023. A quantitative assessment of the status of benthic communities on U.S. Atlantic coral reefs using a novel standardized approach. *Bull Mar Sci.* 99:1-24. <https://doi.org/10.5343/bms.2022.0025>
- Waddell, J. 2017. Florida Reef Resilience Program Disturbance Response Monitoring quick look report summary. National Ocean Service (NOS), National Oceanic and Atmospheric Administration (NOAA), Coral Reef Conservation Program (CRCP), The Nature Conservancy (TNC). <https://repository.library.noaa.gov/view/noaa/15575>
- Walton CJ, Hayes NK, Gilliam DS. 2018. Impacts of a regional, multi-year, multi-species coral disease outbreak in Southeast Florida. *Front Mar Sci.* 5. <https://doi.org/10.3389/fmars.2018.00323>
- Williams DE, Miller MW, Bright A, Pausch R, Valdiva A. 2017. Thermal stress exposure, bleaching response, and mortality in the threatened coral *Acropora palmata*. *Mar Poll Bull.* 124:1. <https://doi.org/10.1016/j.marpolbul.2017.07.001>
- Williams DE, Nedimyer K, Miller MW. 2020. Genotypic inventory of *Acropora palmata* (elkhorn coral) populations in south Florida, Protected Resources and Biodiversity Division Report: NOAA/SEFSC/PRBD-2020-01, NOAA NMFS Southeast Fisheries Science Center.

TABLES

Table 1. Regions, subregions, habitat types, and depths that defined analysis strata (AS) in the South Florida coral reef survey. Stratum-specific areas (A_h), total possible primary units (N_h), and sampled primary units (n_h) are listed.

AS Strata Code	Region/Subregion	Habitat	n_h	N_h	A_h (km^2)
Dry Tortugas					
AS01	Tortugas Bank (BANK)	Contiguous Reef (CONT), High Relief (HR), all depths	32	1766	17.66
AS02	National Park (DTNP)	CONT, HR, all depths	5	507	5.07
AS03	DTNP	CONT, Medium Relief (MR), all depths	41	1479	14.79
AS04	BANK	CONT, Low Relief (LR), all depths	10	9788	97.88
AS05	DTNP	CONT, LR, all depths	31	7391	73.91
AS06	BANK	Isolated Patch Reef (ISOL), HR, all depths	5	260	2.60
AS07	DTNP	ISOL, HR, all depths	22	510	5.10
AS08	BANK	ISOL, MR, all depths	10	1447	14.47
AS09	DTNP	ISOL, MR, all depths	12	3305	33.05
AS10	BANK	ISOL, LR, all depths	7	268	2.68
AS11	DTNP	ISOL, LR, all depths	15	2767	27.67
AS12	DTNP	Spur and Groove Reef (SPGR), HR, all depths	11	359	3.59
AS13	DTNP	SPGR_LR, all depths	3	1251	12.51
Florida Keys					
AS14	all Subregions	Inshore Patch Reef (IPR), all relief types and depths	15	1684	16.84
AS15	all Subregions	Mid-channel Patch Reef (MPR), all relief types and depths	76	15192	151.92
AS16	all Subregions	Offshore Patch Reef (OPR), all relief types and depths	53	9159	91.59
AS17	all Subregions	Fore reef (FRF), Low relief Rubble (RUBB_LR), all depths	30	1248	12.48
AS18	all Subregions	FRF, SPGR, HR, all depths	83	1170	11.70
AS19	all Subregions	FRF, LR, Shallow (S) (0m - 6m)	18	3238	32.38
AS20	all Subregions	FRF, LR, Mid-Shallow (MS) ($\geq 6\text{m} - 12\text{m}$)	100	17807	178.07
AS21	all Subregions	FRF, LR, Mid-Deep (MD) ($\geq 12\text{m} - 18\text{m}$)	33	7951	79.51
AS22	all Subregions	FRF, LR, Deep (D) ($\geq 18\text{m} - 33\text{m}$)	7	9593	95.93
Southeast Florida					
AS23	Broward-Miami (BM)	Nearshore Ridge (NEAR), LR	20	5568	55.68
AS24	BM	NEAR, HR	5	160	1.60
AS25	BM	Linear Reef Inner (INNER), LR	10	1204	12.04

AS26	BM	INNER, HR	11	605	6.05
AS27	BM	Linear Reef Middle (MIDR), LR	48	1263	12.63
AS28	BM	MIDR, HR	25	428	4.28
AS29	BM	Linear Reef Outer (OFFR), LR	5	630	6.30
AS30	BM	OFFR, HR	15	1227	12.27

Table 2. Regions, subregions, habitat types, depths, and spatial management areas that defined condensed field strata (FS-C) in the South Florida coral reef survey. Stratum-specific total possible primary units (N_h) and sampled primary units (n_h) are listed. Respective habitat use strata (HUS) for *Orbicella faveolata*, *Orbicella franksi*, *Orbicella annularis*, and *Acropora cervicornis* were delineated by combining FS with similar means and variances of colony density (e.g., Fig. 3).

Field Strata, Condensed (FS-C)					Species -Specific Habitat Use Strata (HUS)				
FS-C Strata Code	Region/Subregion	Habitat	Spatial management	n	N _h	<i>O. faveolata</i>	<i>O. franksi</i>	<i>O. annularis</i>	<i>A. cervicornis</i>
Dry Tortugas									
AS01/AS02_FSC01	Tortugas Bank (BANK)/National Park (DTNP)	Contiguous Reef, High Relief (CONT_HR), all depths	Open (OPN) + MPA	37	2273	1	1	1	1
AS03_FSC02	DTNP	CONT, Medium Relief (CONT_MR), all depths	OPN	21	953	1	1	3	4
AS03_FSC03	DTNP	CONT_MR	MPA	20	526	1	1	1	4
AS04_FSC04	BANK	CONT, Low Relief (CONT_LR), all depths	OPN + MPA	10	9788	2	2	2	2
AS05_FSC05	DTNP	CONT_LR, Shallow (S) (< 12m)	Open	9	1590	2	2	2	3
AS05_FSC06	DTNP	CONT_LR_S	MPA	4	883	2	2	2	2
AS05_FSC07	DTNP	CONT_LR, Deep (D) (12m - 33m)	OPN + MPA	18	4918	2	3	1	2
AS06_FSC08	BANK	Isolated Patch Reef, High Relief (ISOL_HR), all depths	OPN + MPA	5	260	1	1	1	1
AS07_FSC09	DTNP	ISOL_HR	OPN + MPA	22	510	1	1	1	1
AS08_FSC10	BANK	ISOL, Medium Relief (MR), Shallow (S) (< 22m)	OPN + MPA	3	336	3	1	1	1
AS08_FSC11	BANK	ISOL_MR, Deep (D) (≥ 22m - 33m)	OPN + MPA	7	1111	4	1	1	1
AS09_FSC12	DTNP	ISOL_MR, Shallow (< 12m)	OPN + MPA	6	1724	5	1	1	4
AS09_FSC13	DTNP	ISOL_MR, Deep (≥ 12m - 33m)	OPN + MPA	6	1581	6	1	1	4
AS10_FSC14	BANK	ISOL, Low Relief (LR), all depths	OPN + MPA	7	268	2	2	2	2
AS11_FSC15	DTNP	ISOL_LR	OPN + MPA	15	2767	2	2	2	2
AS12_FSC16	DTNP	Spur and Groove Reef (SPGR), High Relief (HR), all depths	OPN + MPA	11	359	1	1	4	1
AS13_FSC17	DTNP	SPGR, Low Relief (LR), all depths	OPN + MPA	3	1251	2	2	2	2

Field Strata, Condensed (FS-C)						Species -Specific Habitat Use Strata (HUS)			
FS-C Strata Code	Region/Subregion	Habitat	Spatial management	n	N _h	<i>O. faveolata</i>	<i>O. franksi</i>	<i>O. annularis</i>	<i>A. cervicornis</i>
Florida Keys									
AS14_FSC18	Lower Keys (L)	Inshore Patch Reef (IPR) , all relief types and depths	OPN + MPA	11	701	7	4	5	5
AS14_FSC19	Middle Keys (M)	IPR	OPN + MPA	1	558	7	4	6	5
AS14_FSC20	Biscayne/Upper Keys (B/U)	IPR	Open (OPN) + MPA	3	425	8	5	6	5
AS15_FSC21	L	Mid-channel Patch Reef (MPR), all relief types and depths	Open (OPN)	25	4952	9	4	7	6
AS15_FSC22	L	MPR	MPA	6	277	10	4	8	6
AS15_FSC23	M	MPR	OPN + MPA	18	1909	11	4	9	7
AS15_FSC24	U	MPR	OPN + MPA	12	2793	11	5	9	7
AS15_FSC25	B	MPR	OPN + MPA	15	5261	12	5	9	8
AS16_FSC26	L	Offshore Patch Reef (OPR), all relief types and depths	OPN + MPA	12	2463	13	4	10	9
AS16_FSC27	M	OPR	OPN + MPA	6	979	14	4	11	9
AS16_FSC28	U	OPR	OPN	16	4346	15	5	12	10
AS16_FSC29	U	OPR	MPA	2	56	16	5	12	10
AS16_FSC30	B	OPR	OPN + MPA	6	910	17	5	12	10
AS16_FSC31	U	Inner Line Reef Tract (IRT), all relief types and depths	OPN	5	346	15	5	12	11
AS16_FSC32	U	IRT	MPA	6	59	16	5	12	11
AS17_FSC33	M/L	Fore reef (FRF), Low relief Rubble (RUBB_LR), all depths	OPN + MPA	23	862	18	6	15	15
AS17_FSC34	B/U	FRF_RUBB_LR	OPN + MPA	7	386	18	6	16	15
AS18_FSC35	L	FRF, High Relief Spur and Groove (SPGR_HR), all depths	OPN + MPA	37	461	19	7	13	12
AS18_FSC36	M	FRF_SPGR_HR	OPN + MPA	26	106	19	7	14	13
AS18_FSC37	U	FRF_SPGR_HR	OPN	6	186	20	8	14	14
AS18_FSC38	U	FRF_SPGR_HR	MPA	6	286	21	8	14	14
AS18_FSC39	B	FRF_SPGR_HR	OPN + MPA	8	131	22	8	14	14

Field Strata, Condensed (FS-C)						Species -Specific Habitat Use Strata (HUS)			
FS-C Strata Code	Region/Subregion	Habitat	Spatial management	n	N _h	<i>O. faveolata</i>	<i>O. franksi</i>	<i>O. annularis</i>	<i>A. cervicornis</i>
Florida Keys									
AS19_FSC40	M/L	FRF, Low Relief (LR), Shallow (S) (0m - 6m)	OPN + MPA	9	986	23	9	15	16
AS19_FSC41	B/U	FRF_LR_S	OPN + MPA	9	2252	24	10	16	17
AS20_FSC42	L	FRF_LR, Mid-Shallow (MS) (≥ 6m - 12m)	OPN + MPA	33	4379	25	9	15	18
AS20_FSC43	M	FRF_LR_MS	OPN	25	7199	25	9	15	19
AS20_FSC44	M	FRF_LR_MS	MPA	19	202	25	9	15	20
AS20_FSC45	U	FRF_LR_MS	OPN + MPA	13	3304	26	10	16	21
AS20_FSC46	B	FRF_LR_MS	OPN + MPA	10	2723	26	10	16	22
AS21_FSC47	M/L	FRF_LR, Mid-Deep (MD) (≥ 12m - 18m)	OPN + MPA	20	5483	25	9	15	23
AS21_FSC48	B/U	FRF_LR_MD	OPN + MPA	13	4922	26	10	16	24
AS22_FSC49	M/L	FRF_LR, Deep (D) (≥ 18m - 33m)	OPN	6	6544	27	11	15	23
AS22_FSC50	B/U	FRF_LR_D	OPN	1	3049	27	11	16	24
Southeast Florida									
AS23_FSC51	Broward-Miami (BM)	Nearshore Ridge (NEAR), Low Rugosity (LR)	Open (OPN)	20	5568	28	12	17	25
AS24_FSC52	BM	NEAR, High Rugosity (HR)	OPN	5	160	28	12	17	26
AS25_FSC53	BM	Linear Reef Inner (INNER), Low Rugosity (LR)	OPN	10	1204	28	12	17	26
AS26_FSC54	BM	INNER, High Rugosity (HR)	OPN	11	605	28	12	18	26
AS27/AS28/AS29/ AS30_FSC55	BM	Linear Reef Middle, Low and High Rugosity (MIDR) and Linear Reef Outer, Low and High Rugosity (OFFR)	OPN	93	3548	28	12	17	26

Table 3. The projected sample size n^* to achieve a desired level of precision (%CV) for eight species and two stratification schemes: (A) analysis (30 total strata) and (B) composite species habitat use (55 total strata). Gray shading represents the number of samples at or below the sampling effort of the 2016 NCRMP/DRM survey ($n=451$). Note that the practical minimum sample size is twice the number of strata: $n=60$ for the AS scheme (A), and $n=110$ for the composite species HUS scheme (B).

(A)

Species	Mean density	Mean proportion occurrence	n*							
			4%	5%	7.50%	10%	15%	20%	30%	40%
<i>S. siderea</i>	1.2816	0.8520	393	252	112	63	28	16	7	4
<i>M. cavernosa</i>	0.2310	0.6155	866	556	248	140	62	35	16	9
<i>O. faveolata</i>	0.0752	0.2602	1651	1068	480	271	121	68	30	17
<i>O. franksi</i>	0.0379	0.1395	1287	838	379	214	96	54	24	14
<i>A. cervicornis</i>	0.0196	0.0423	9195	6269	2978	1717	777	440	196	111
<i>O. annularis</i>	0.0171	0.0462	2637	1802	858	495	224	127	57	32
<i>M. ferox</i>	0.0011	0.0100	14624	10761	5613	3361	1566	896	403	228
<i>A. palmata</i>	0.0011	0.0035	8811	7381	4721	3138	1602	951	440	251

(B)

Species	Mean density	Mean proportion occurrence	n*							
			4%	5%	7.50%	10%	15%	20%	30%	40%
<i>S. siderea</i>	1.2319	0.8535	250	160	71	40	18	10	4	3
<i>M. cavernosa</i>	0.2214	0.6075	674	433	193	109	48	27	12	7
<i>O. faveolata</i>	0.0741	0.2600	1196	772	346	195	87	49	22	12
<i>O. franksi</i>	0.0376	0.1420	993	646	292	165	74	42	18	10
<i>A. cervicornis</i>	0.0205	0.0425	4475	3040	1438	827	374	212	94	53
<i>O. annularis</i>	0.0138	0.0395	2515	1724	824	476	216	122	55	31
<i>M. ferox</i>	0.0013	0.0110	6130	4453	2284	1358	629	359	161	91
<i>A. palmata</i>	0.0005	0.0011	2228	2000	1475	1078	610	379	182	106

Table 4. Projected number of samples (n^*) required to achieve a 15% CV for mean density at the full domain (southern Florida) and region level for two abundant species and six ESA-listed species using the composite habitat use stratification. The species are listed in order of abundance from left to right: *S. siderea* (SSID), *M. cavernosa* (MCAV), *O. faveolata* (OFAV), *O. franksi* (OFRA), *A. cervicornis* (ACER), *O. annularis* (OANN), *M. ferox* (MFER), and *A. palmata* (APAL).

Sampling frame	NCRMP/DRM n	$n^*_{15\%}$								Average
		SSID	MCAV	OFAV	OFRA	ACER	OANN	MFER	APAL	
Southern Florida	451	18	48	87	74	374	216	629	610	257
<i>Regions</i>										
Broward-Miami	98	80	149	247	575	681	584	3172	3172	1083
Florida Keys	185	17	46	78	242	301	204	826	467	273
Dry Tortugas	168	24	42	149	48	293	1101	418	3889	746
<i>Region total</i>	451	121	237	474	865	1275	1889	4416	7528	2101

FIGURE CAPTIONS

Figure 1. Location of 758 Primary Sample Units (PSUs) within the southern Florida study area for NCRMP and DRM surveys in 2013-2016. Sampling encompassed the Broward-Miami subregion in Southeast Florida, four subregions in the Florida Keys (Biscayne, Upper, Middle, Lower Keys), and two subregions in the Dry Tortugas (Dry Tortugas National Park, Tortugas Bank). The Marquesas subregion was not sampled due to limited bathymetry and benthic habitat data.

Figure 2. Relationships between coefficient of variation (CV) of mean density and predicted sample size n^* for six ESA-listed coral species and different stratification schemes. Two schemes, analysis strata (AS, gray line) and simple random (SRS, dashed line), were evaluated for all six species. An additional scheme, species-specific habitat use strata (HUS, dashed line), was evaluated for four species (A-D). The projected n^* to achieve a 15% CV is denoted.

Figure 3. Construction of habitat use strata (HUS), illustrated for *O. faveolata* in the Florida Keys region. Panels show the spatial pattern of density at progressively smaller spatial scales: (A) cross-shelf patch reef habitats, (B) subregions within Mid-channel patch reefs, and (C) management zones within Mid-channel patch reefs in the Lower Keys subregion. Dashed lines are the area-weighted mean density of individual habitats in each panel. Vertical lines denote delineated HUS based on similarities or differences in the spatial density pattern. Nonparametric tests for the final HUS groupings: Wilcoxon (W) test for differences in means; Conover (C) test for differences in means and variances. Habitat, subregion, and spatial management type codes are defined in Table 2.

FIGURES

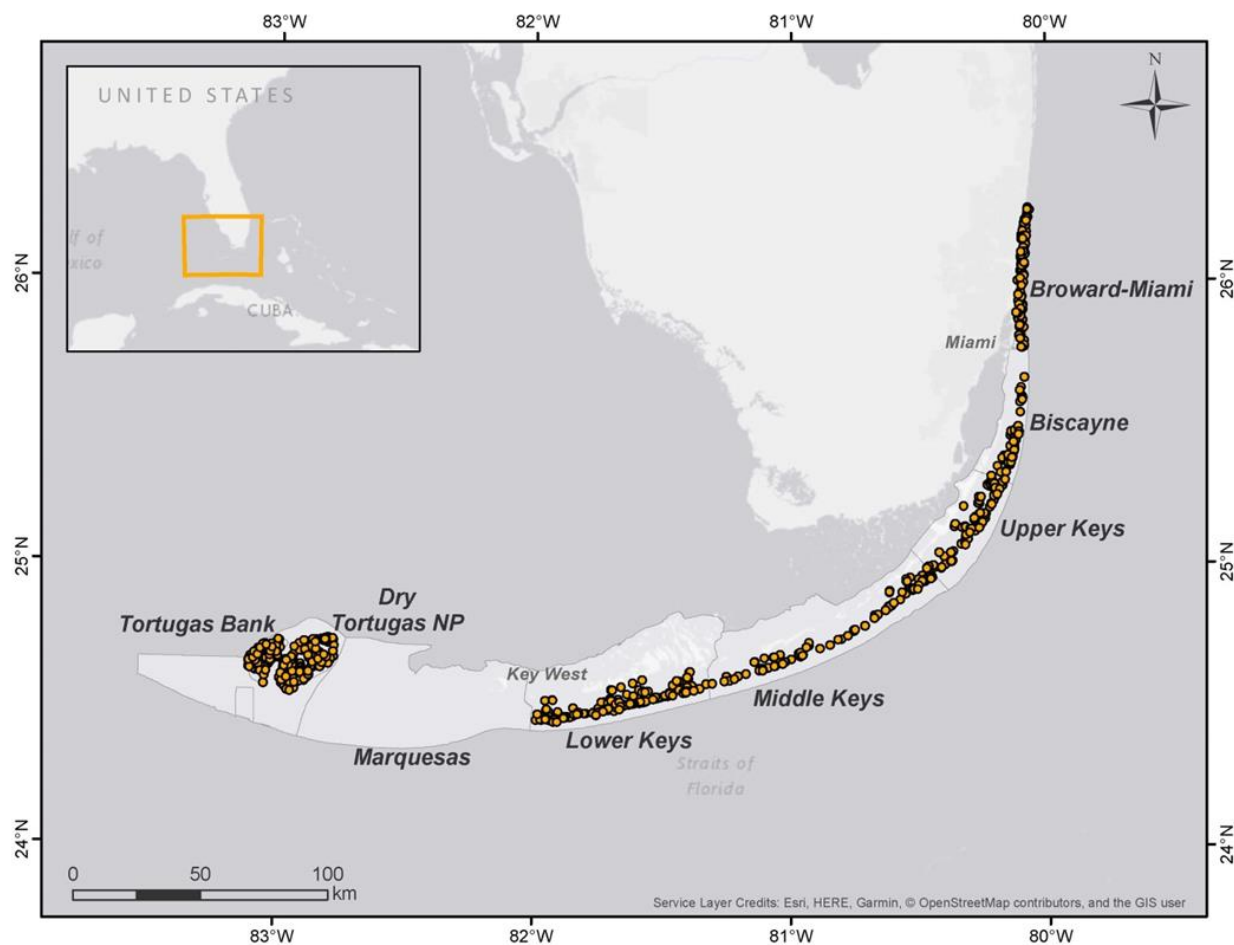


Figure 1.

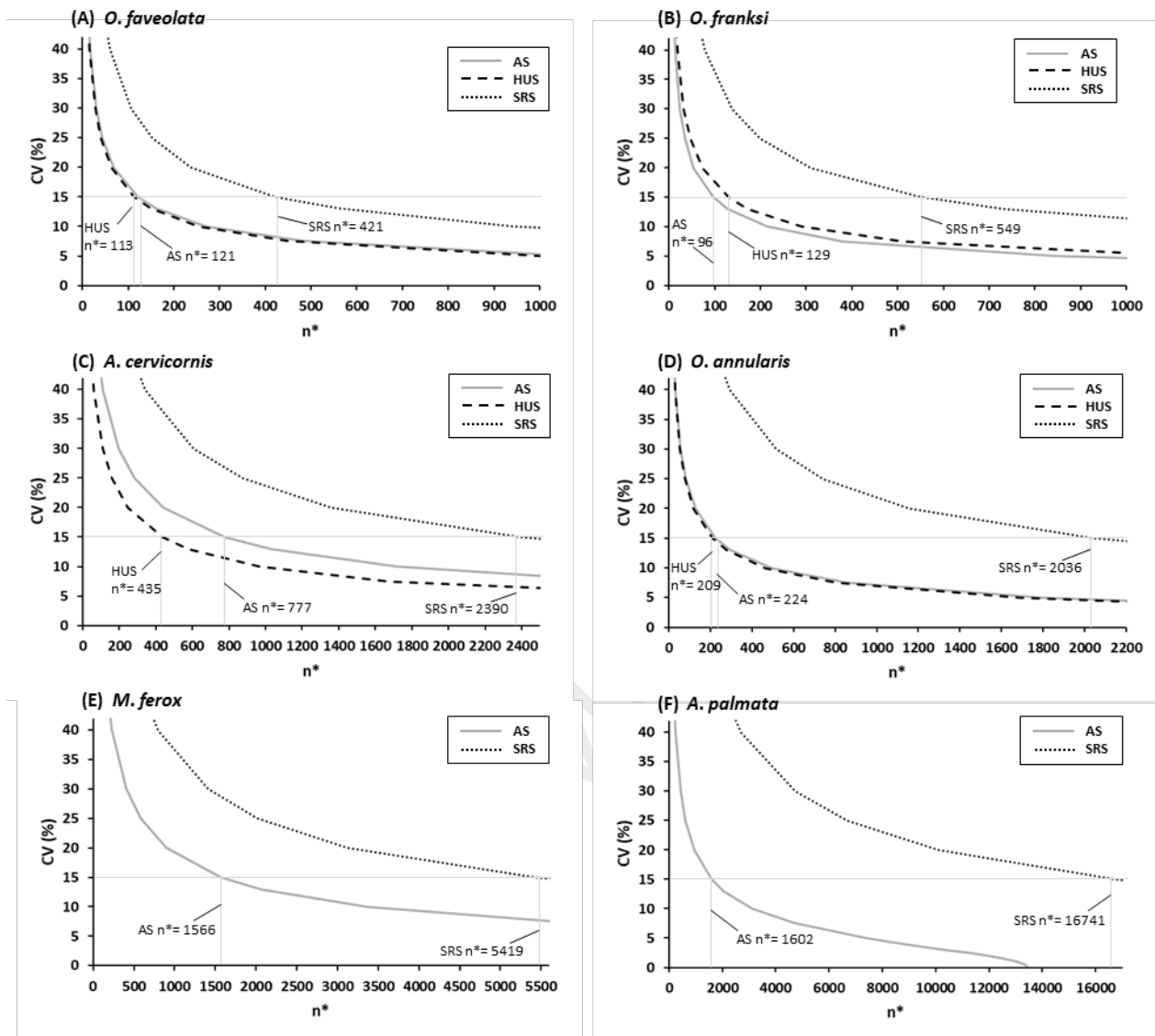
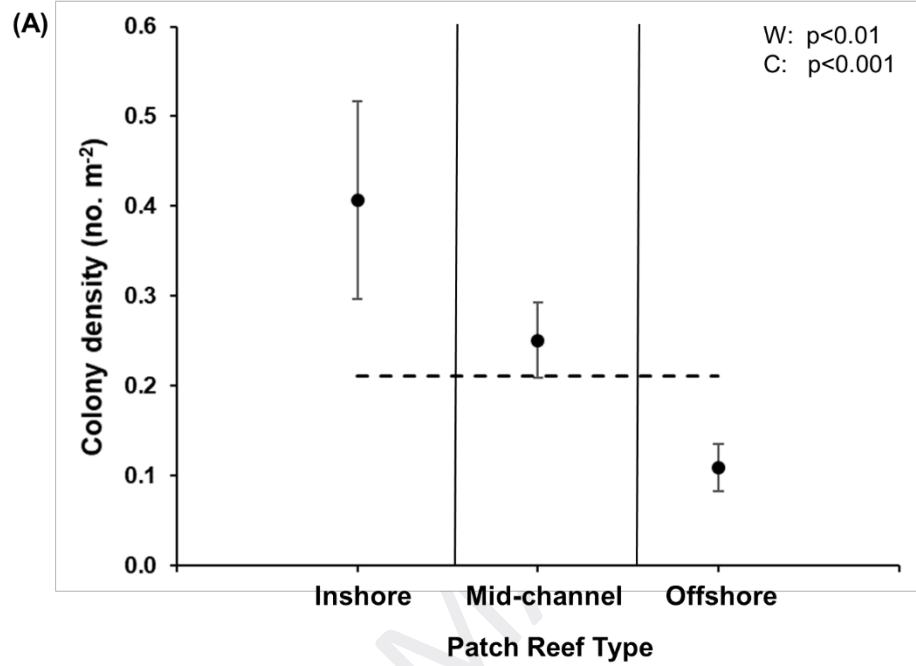


Figure 2.



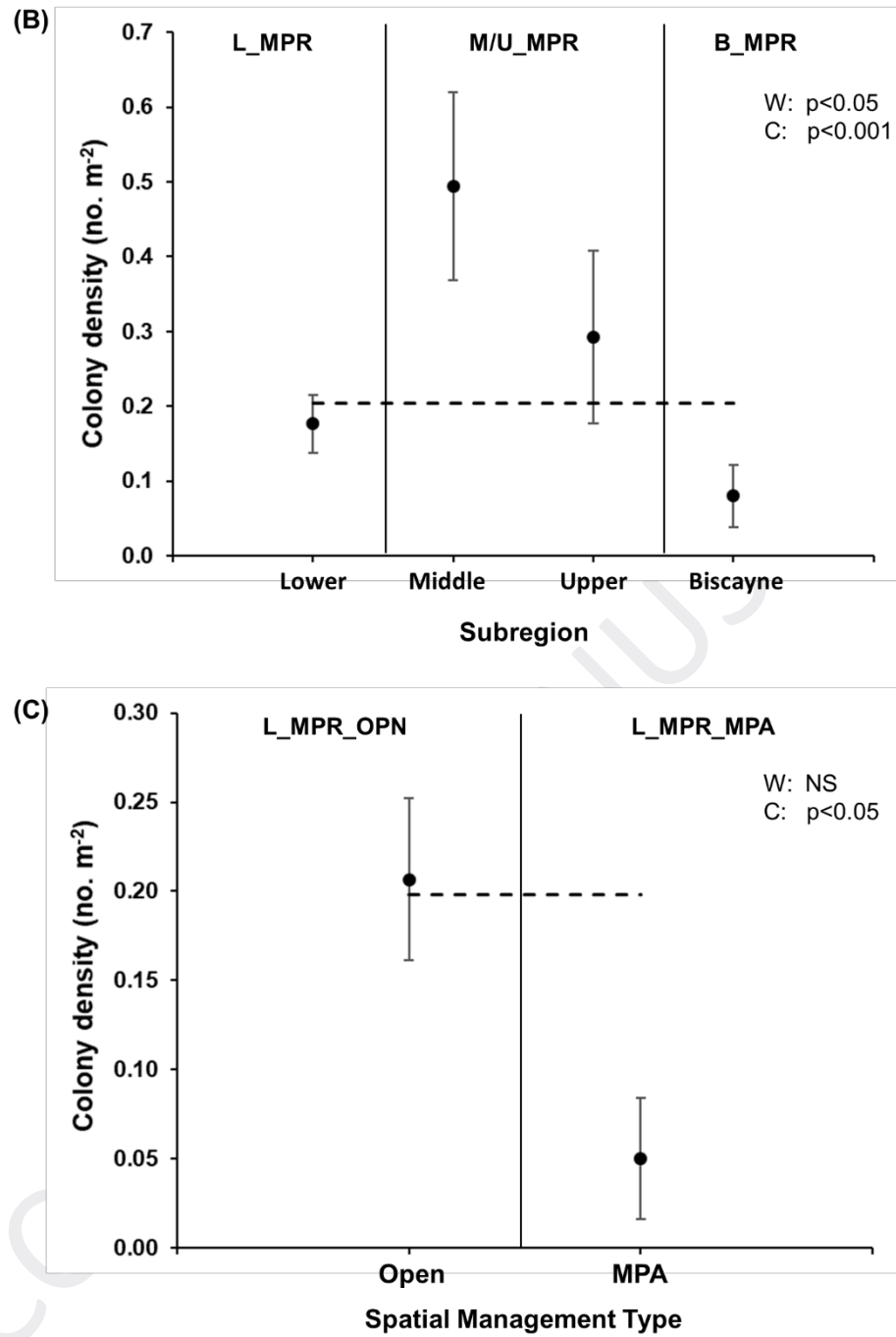


Figure 3.